

The Putative Interactome of *Escherichia coli* Glutaredoxin 2 as revealed by Affinity Chromatography

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Introduction. Glutaredoxin 2 (Grx2) contributes more than 80 % of glutathione (GSH) mediated redox activity in cellular extracts and protects cells from general oxidative damage (formation of carbonyls^{1,2}). The specific substrates of Grx2 however, are unknown. To this aim, immobilized monothiol/athiol Grx2 mutants were used in affinity chromatography with cellular lysates from *Escherichia coli* (*E. coli*). The athiol Grx2 (Grx2 C9S C12S) served as a bait for proteins interacting in a non-thiol manner while the monothiol Grx2 C12S was used to trap dithiol substrates of Grx2. In addition, lysates from *E. coli* null mutants for *grxB*, encoding Grx2, were compared to those of the wild type. All proteomic analyses were performed by Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS) followed by bioinformatics and gene ontology evaluations.

Methodology. Overexpression and purification of *E. coli* Grx2 C9S C12S and Grx2 C12S mutants. Each protein (approximately 6 mg) was immobilized on Affi-Gel 10 beads in chromatographic columns. *E. coli* cells were grown in LB-medium and collected during both exponential and stationary phases. Cell lysates were prepared and chromatographed through columns with of the immobilized Grx2 mutants. Empty resin served as control. Bound proteins were eluted with salt (KCl, step gradients), CH₃COOH/HCOOH and finally DTT. The eluted proteins were analyzed by LC-MS/MS. All cromatographies were performed in triplicates. Furthermore, *E. coli* wild type strain and the null mutant for *grxB* were grown and harvested in both growth phases. Their proteome was analyzed by LC-MS/MS.

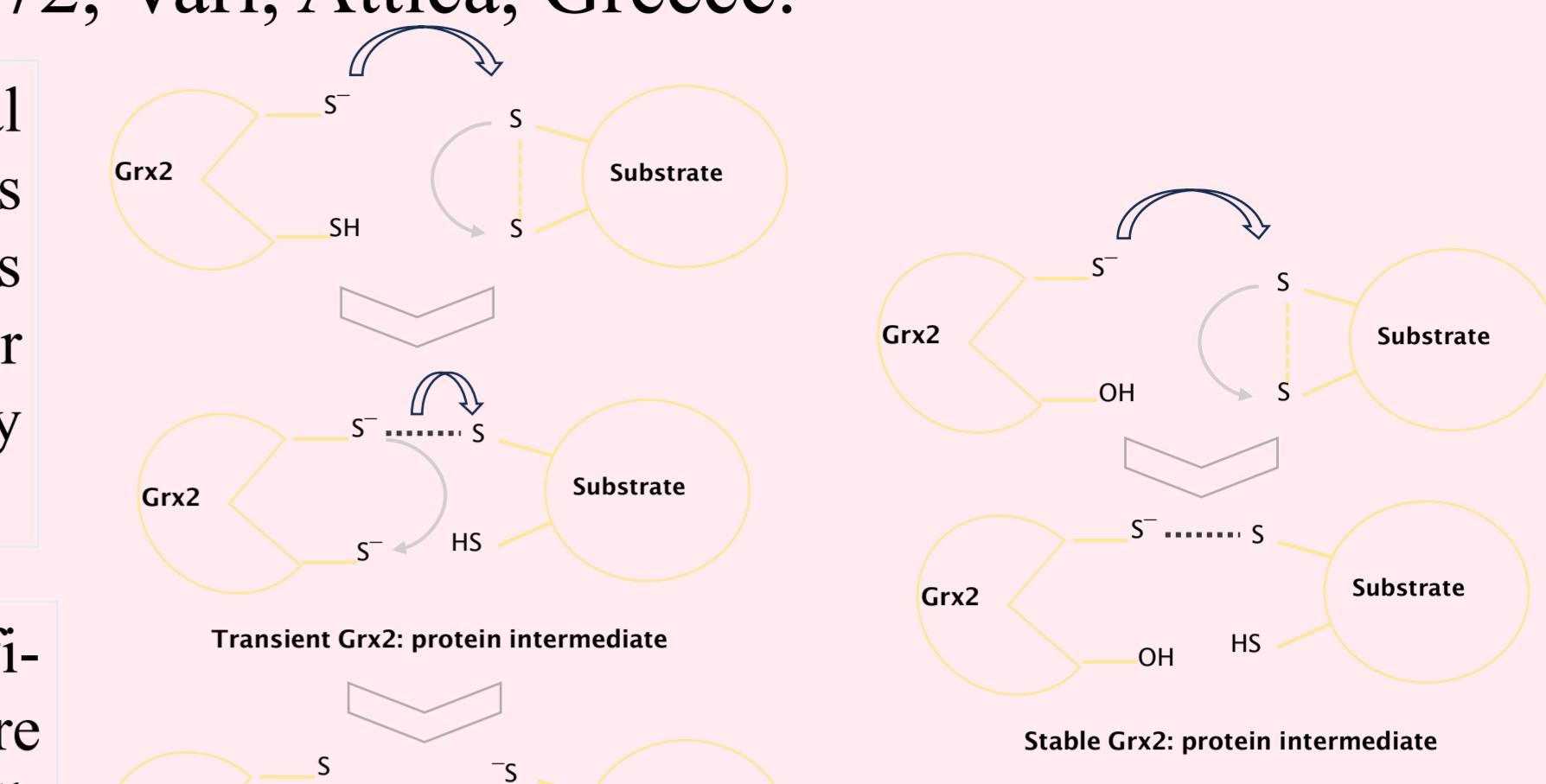


Figure 1. Catalytic mechanism and monothiol Grx substrate trap.

Results

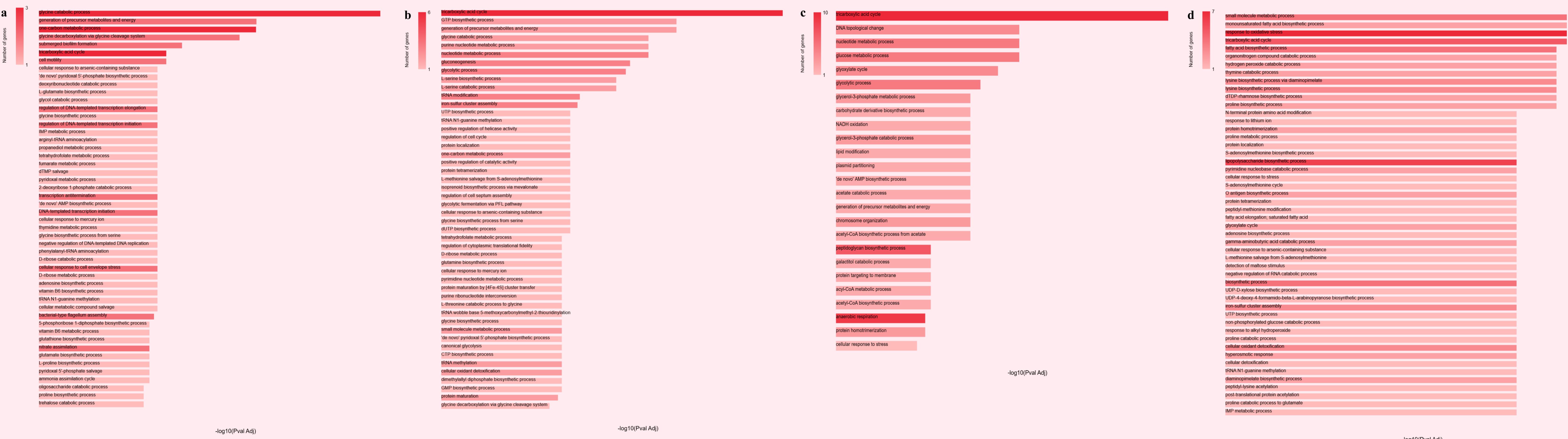


Figure 2 Gene Ontology annotation of the statistically significant proteins, derived from affinity chromatography, performed in GeneCodis4. Ontologies were retrieved from the databases Biological process and Molecular function. Exponential phase acidic elution (a) and DTT elution (b). Stationary phase acidic elution (c) and DTT elution (d).

Table 1. Top 50 potential interactors of each growth phase, in acidic and DTT elutions, were selected for docking experiments.

Docking simulation					
Acidic elution	Exponential Phase		Stationary Phase		Total 63% agreement
	52% (13 complexes)	44% (11 complexes)	84% (21 complexes)	72% (18 complexes)	
DTT elution					

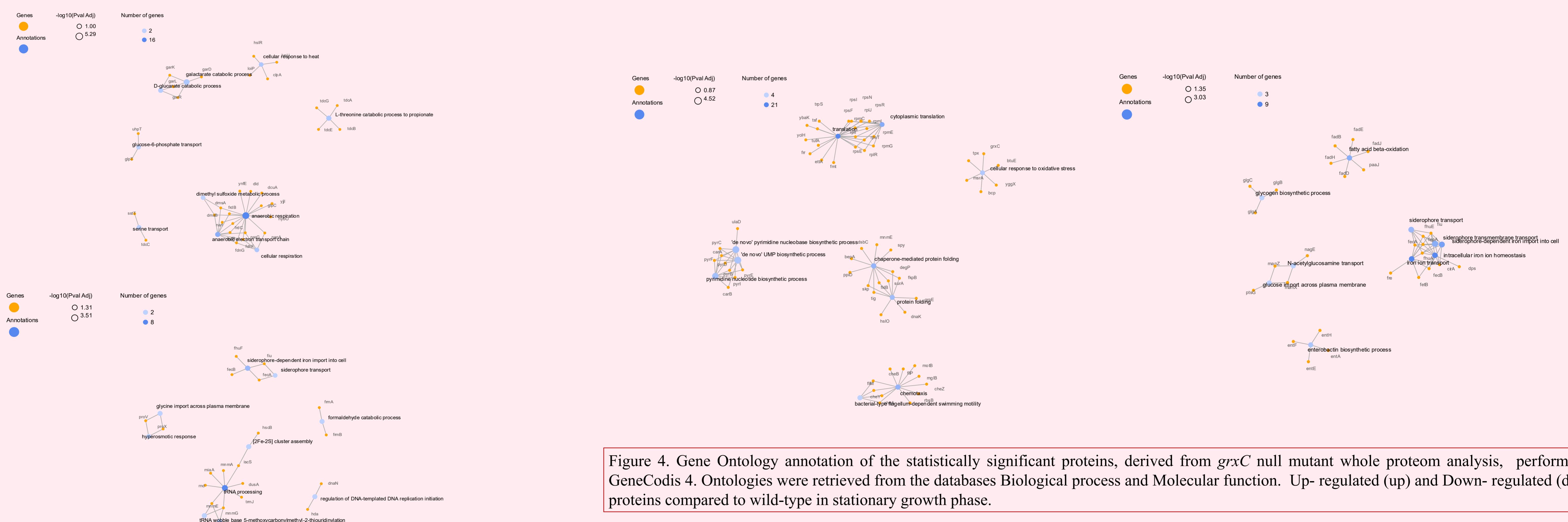

 Figure 4. Gene Ontology annotation of the statistically significant proteins, derived from *grxC* null mutant whole proteom analysis, performed in GeneCodis 4. Ontologies were retrieved from the databases Biological process and Molecular function. Up-regulated (up) and Down-regulated (down) proteins compared to wild-type in stationary growth phase.

 Figure 3. Gene Ontology annotation of the statistically significant proteins, derived from *grxC* null mutant whole proteom analysis, performed in GeneCodis 4. Ontologies were retrieved from the databases Biological process and Molecular function. Up-regulated (up) and Down-regulated (down) proteins compared to wild-type in exponential growth phase.

Conclusions

1. Affinity chromatography experiments revealed that Grx2-interacting proteins may be involved tricarboxylic acid cycle, response to oxidative stress, peptidoglycane biosynthesis, iron-sulfur cluster assembly and regulation of cell cycle.
2. *grxB*⁻-wild type whole proteome comparisons showed significant changes in the levels of proteins involved in anaerobic respiration, tRNA processing, iron ion homeostasis, protein folding and chemotaxis.
3. Grx2 appears as a multifunctional protein involved in many more biological pathways than its known general antioxidant function.

References

1. Alexios Vlamis-Gardikas . (2008). The multiple functions of the thiol-based electron flow pathways of *Escherichia coli*: External concepts revisited. *Biochim Biophys Acta* 1780(11):1170-200.
2. Ogata F. T., Branco V., Vale F. F., Coppo, L. (2021). Glutaredoxin: Discovery, redox defense and much more. *Redox Biology*, 43, 101975.

Acknowledgements

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